

## DICLOFENAC SODIUM (GP 45840, VOLTAREN), A POTENT INHIBITOR OF PROSTAGLANDIN SYNTHETASE

EDMOND C. KU, JONG M. WASVARY and WILLIAM D. CASH

Research Department, Pharmaceuticals Division, CIBA-GEIGY Corp., Ardsley, N.Y. 10502, U.S.A.

(Received 17 May 1974; accepted 16 August 1974)

**Abstract**—Diclofenac sodium, the sodium salt of *o*-(2,6-dichlorophenylamino)-phenylacetic acid (GP 45840, Voltaren), is a potent inhibitor of sheep seminal vesicle prostaglandin synthetase *in vitro*. Like indomethacin, aspirin and certain other nonsteroidal anti-inflammatory drugs, it acts as a competitive and irreversible inhibitor. It is suggested that diclofenac sodium exerts most of its pharmacological effects via inhibition of prostaglandin synthetase.

Diclofenac sodium, the sodium salt of *o*-(2,6-dichlorophenylamino)-phenylacetic acid (GP 45840, Voltaren), is a new, clinically effective anti-inflammatory agent [1–6]. In animal models of acute and chronic inflammation, it is distinctly more active than phenylbutazone and comparable in potency to indomethacin [1]. Its therapeutic index (the ratio of acutely toxic doses in mice or rats to effective doses in anti-inflammatory models) is higher than that of phenylbutazone or indomethacin [1]. It exhibits analgesic and antipyretic activity in appropriate animal tests and prevents bradykinin-induced bronchoconstriction in guinea pigs [1]. It also prevents platelet aggregation under various conditions [7, 8].

The pharmacological effects of diclofenac sodium are consistent with a mechanism of action involving inhibition of prostaglandin (PG) synthesis, a mechanism that appears to explain most of the biological effects of indomethacin, aspirin and other nonsteroidal anti-inflammatory agents [9–15]. It was of interest, therefore, to evaluate diclofenac sodium directly for its ability to inhibit PG synthetase. The results of studies *in vitro* with sheep seminal vesicle PG synthetase are presented in this report.

### MATERIALS AND METHODS

Lyophilized sheep seminal vesicle microsomes [16] served as the source of PG synthetase. The enzyme was assayed by a modification of the procedure described by Takeguchi *et al.* [16]. Enzyme preparation (10–20  $\mu$ g/ml),  $^{14}$ C-arachidonic acid (0.4 to 6  $\mu$ M, obtained from Applied Science Labs, Inc., sp. act. 14–58 mCi/m-mole), epinephrine (1 mM), glutathione (1 mM) and the compound being tested were incubated at 25° in 0.1 M Tris buffer (pH 8.3, total volume usually 1 ml). Arachidonic acid was first dissolved in ethanol; the final concentration of ethanol was kept constant in each exper-

iment and did not exceed 5% (v/v). This level of ethanol did not affect the enzyme assay. Compounds being tested were dissolved in ethanol or directly in buffer, in some cases with the aid of NaOH. Care was taken that the final pH of the incubation medium did not deviate from pH 8.3.

After an appropriate incubation period (10 min unless otherwise indicated), the enzyme reaction was stopped by the addition of one drop of concentrated HCl/ml of incubation medium. The acidified mixture was saturated with sodium sulfate and extracted with two 5-ml portions of ethyl acetate containing 7  $\mu$ M arachidonic acid. The latter was added as a carrier and also to facilitate the subsequent thin-layer chromatographic separation of labeled compounds. The combined extracts were evaporated to dryness. The residue was dissolved in acetone and chromatographed on thin-layer plates in the solvent system described by Tomlinson *et al.* [17]. The plates were scanned with a Varian aerograph radio scanner; the radioactive zones corresponding to PGE<sub>2</sub> (the predominant product) were scraped off, transferred to liquid scintillation vials and counted.

### RESULTS AND DISCUSSION

As shown in Fig. 1, concentrations of diclofenac sodium as low as 1–4  $\mu$ M interfered markedly with the synthesis of PGE<sub>2</sub> from arachidonic acid. The rate of synthesis was depressed initially and remained so for the duration of the experiment. This is indicative of irreversible inhibition as occurs with indomethacin, aspirin or 5,8,11,14-eicosatetraynoic acid [18–21]. As is evident from the data presented in Fig. 2, diclofenac sodium inhibited the enzyme competitively with respect to substrate, arachidonic acid.

Inhibition constants ( $K_i$  values) for diclofenac sodium and several other PG synthetase inhibitors are presented in Table 1. All of the compounds listed, with

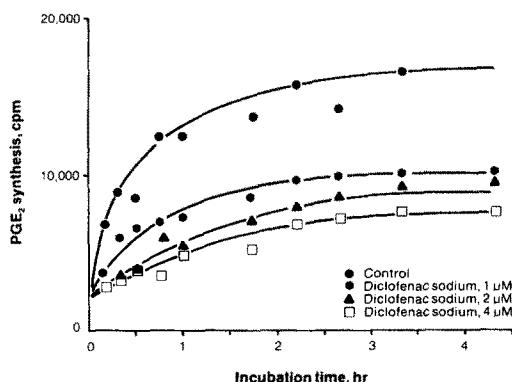


Fig. 1. Effect of diclofenac sodium on PGE<sub>2</sub> synthesis from arachidonic acid. The initial arachidonic acid concentration was 6  $\mu$ M. PGE<sub>2</sub> synthesis is expressed as counts/min/0.5 ml of incubation medium. The results of a typical experiment are shown; each point represents a single measurement. The same pattern of inhibition was consistently noted in additional experiments.

the exception of 2,7-naphthalenediol, behaved as competitive inhibitors in these experiments. The competitive inhibitors acted irreversibly, although to varying extents.\* Thus, with respect to the mode of inhibition, diclofenac sodium resembles indomethacin, aspirin and certain other nonsteroidal anti-inflammatory drugs. The low  $K_i$  of diclofenac sodium indicates that it is among the most potent compounds of the group.

A further point of resemblance between diclofenac sodium and indomethacin is the failure of either agent to cause complete irreversible inhibition of sheep seminal vesicle PG synthetase under our experimental conditions. Concentrations of diclofenac sodium higher than those shown in Fig. 1 did not further

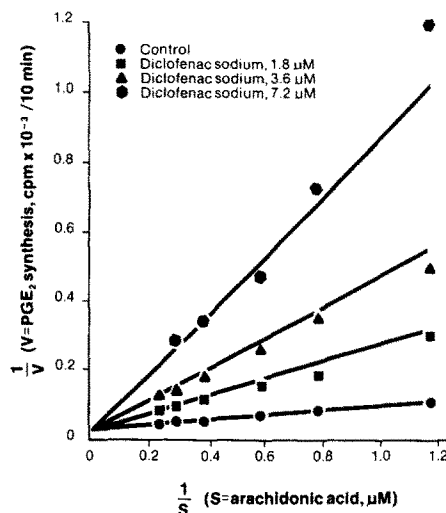


Fig. 2. Double reciprocal plot showing competitive inhibition of PGE<sub>2</sub> synthesis from arachidonic acid by diclofenac sodium. PGE<sub>2</sub> synthesis is expressed as counts/min ( $\times 10^{-3}$ )/0.5 ml of incubation medium during a 10-min incubation period. The results of a typical experiment are shown; each point represents a single measurement. The same pattern of inhibition was noted in two additional experiments.

reduce the yield of PGE<sub>2</sub>. This behavior was exhibited by indomethacin also. However, aspirin was capable of producing complete irreversible inhibition. Published studies with 5,8,11,14-eicosatetraynoic acid suggest that it too causes total irreversible inhibition [20].

A likely explanation for the behavior of diclofenac sodium and the other competitive, irreversible inhibitors is that they induce or enhance inactivation of the enzyme subsequent to formation of the enzyme-inhibitor complex [18–21]. We cannot satisfactorily explain the difference between diclofenac sodium and indomethacin on the one hand, and aspirin and 5,8,11,14-eicosatetraynoic acid on the other, with respect to the

\* Reversibility was assessed by determining the net synthesis of PGE<sub>2</sub> during a 16-hr incubation in the presence of excess arachidonic acid (initial concentration of 2–6  $\mu$ M) and varying inhibitor concentrations (multiples of the  $K_i$ ). Under such conditions, PGE<sub>2</sub> synthesis was markedly reduced by some drugs at concentrations near their  $K_i$  value. For example, indomethacin and diclofenac sodium inhibited synthesis to the extent of about 50 per cent at concentrations 1–2 times the  $K_i$ . On the other hand, concentrations of the potent inhibitor Su-21524 [19] up to 20 times the  $K_i$  did not affect the net production of PGE<sub>2</sub>. We have, therefore, classified the latter compound as a reversible inhibitor and the former as irreversible inhibitors. Some drugs fall between the two extremes. For example, concentrations of phenylbutazone and oxyphenbutazone 7–10 times the  $K_i$  are required to inhibit net PGE<sub>2</sub> synthesis to the extent of 50 per cent under the specified conditions. They are only weakly irreversible in comparison to indomethacin or diclofenac sodium; yet they are not reversible in comparison to Su-21524. Oxyphenbutazone has previously been characterized as a reversible inhibitor in a study under somewhat different experimental conditions [18, 23]. Indomethacin behaved as an irreversible inhibitor in the same study; phenylbutazone was not evaluated.

Table 1.  $K_i$  values for diclofenac sodium and other inhibitors of PGE<sub>2</sub> synthesis\*

Inhibitor	$K_i$ ( $\mu$ M)
Diclofenac sodium	2.4
Meclofenamic acid	2.6
Mefenamic acid	3.2
2,7-Naphthalenediol	3.4
5,8,11,14-Eicosatetraynoic acid	4.9
Indomethacin	6.5
Oxyphenbutazone	730
Phenylbutazone	98
Aspirin	5500

\* These values were determined at substrate concentrations ranging from 0.4 to 5  $\mu$ M and a constant inhibitor concentration [22]. Each value represents the mean of 2–4 separate determinations.

extent of enzyme inactivation. It is conceivable that the crude enzyme system contains multiple forms of PG synthetase with different susceptibilities to inactivation when bound to particular inhibitors.

In summary, diclofenac sodium is a potent inhibitor of sheep seminal vesicle PG synthetase *in vitro*. Like indomethacin, aspirin and certain other nonsteroidal anti-inflammatory drugs, it acts as a competitive and irreversible inhibitor. In view of these properties and the mounting evidence that inhibition of PG synthesis is the main mechanism of action of nonsteroidal anti-inflammatory agents [9–15], we consider it likely that diclofenac sodium exerts most of its pharmacological effects via inhibition of PG synthetase.\*

#### REFERENCES

1. P. J. Krupp, R. Menassé-Gdynia, A. Sallmann, G. Wilhelm, R. Ziel and R. Jaques, *Experientia* **29**, 450 (1973).
2. P. D. Fowler and S. A. Ghazi, *Excerpta Med., Int. Congress Series* **299**, 115 (1973).
3. S. Sacks, *Excerpta Med., Int. Congress Series* **299**, 116 (1973).
4. G. J. Abrams and L. Solomon, *Excerpta Med., Int. Congress Series* **299**, 116 (1973).
5. M. Takatori, Y. Shiokawa and A. Sakuma, *Excerpta Med., Int. Congress Series* **299**, 116 (1973).
6. S. A. Ghazi and P. D. Fowler, *J. Int. med. Res.* **1**, 591 (1973).
7. S. Renaud and F. Lecompte, *Thromb. Diath. haemorrh.* **24**, 577 (1970).
8. F. Jobin and F. T. Gagnon, *Can. J. Physiol. Pharmac.* **49**, 479 (1971).
9. P. J. Piper and J. R. Vane, *Nature, Lond.* **223**, 29 (1969).
10. J. R. Vane, *Nature New Biol.* **231**, 232 (1971).
11. J. B. Smith and A. L. Willis, *Nature New Biol.* **231**, 235 (1971).
12. H. O. J. Collier and C. Schneider, *Nature New Biol.* **236**, 141 (1972).
13. R. Flower, R. Gryglewski, K. Herbaczynska-Cedro and J. R. Vane, *Nature New Biol.* **238**, 104 (1972).
14. S. H. Ferreira, S. Moncada and J. R. Vane, *Br. J. Pharmac.* **49**, 86 (1973).
15. S. Moncada, S. H. Ferreira and J. R. Vane, *Nature, Lond.* **246**, 217 (1973).
16. C. Takeguchi, K. Kohno and C. J. Sih, *Biochemistry* **10**, 2372 (1971).
17. R. V. Tomlinson, H. J. Ringold, M. C. Qureshi and E. Forchielli, *Biochem. biophys. Res. Commun.* **46**, 552 (1972).
18. W. L. Smith and W. E. M. Lands, *J. biol. Chem.* **246**, 6700 (1971).
19. E. C. Ku and J. M. Wasvary, *Fedn Proc.* **32**, 803 (1973).
20. D. T. Downing, D. G. Ahern and M. Bachta, *Biochem. biophys. Res. Commun.* **40**, 218 (1970).
21. J. Y. Vanderhoek and W. E. M. Lands, *Biochim. biophys. Acta* **296**, 374 (1973).
22. H. Lineweaver and D. Burk, *J. Am. chem. Soc.* **56**, 658 (1934).
23. W. E. M. Lands, P. LeTellier, L. Rome and J. Vanderhoek, *Fedn Proc.* **31**, 476 (1972).

---

\* After this manuscript was submitted, it was brought to our attention that two papers describing diclofenac sodium as a potent inhibitor of bovine seminal vesicle PG synthetase are in press [R. Ziel and P. Krupp, *Int. J. clin. Pharmac. Ther. Toxicol.*, in press; R. Ziel and P. Krupp, in *Temperature Regulation and Drug Action, IInd Symp.*, Paris 1974 (Eds. J. Jacob, P. Lomax and E. Schönbaum). S. Karger, Basel, in press].